

Synthetic studies of thiazoline and thiazolidinecontaining natural products – 1. Phosphorus pentachloride-mediated thiazoline construction reaction

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Abstract

Phosphorus pentachloride effectively mediates the cyclization of *N*-acylcysteamine derivatives giving rise to thiazoline rings. Using this method, sterically hindered thiazoline analogs could be constructed and thus segment A (the left half) of micacocidin, a unique antimycoplasma antibiotic, was synthesized efficiently. © 1999 Elsevier Science Ltd. All rights reserved.

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Mycoplasmas infect a wide range of domestic animals, fowl, insects and plants as well as humans, and the recent finding that mycoplasmas promote proliferation of HIV [1] has attracted a great deal of attention. In 1996, a new antibiotic micacocidin (1), which showed specific and potent activity for mycoplasmas, especially for *Mycoplasma pneumoniae*, was isolated in our laboratory in the form of stable metal complexes from the culture broth of *Pseudomonas* sp. Lot 57-250 [2]. The absolute stereostructure of micacocidin A (2), a Zn complex of 1 was determined by X-ray crystallographic analysis. Its biological activities and the novel structure, comprised of one alkylphenol unit and three characteristic thiazoline and thiazolidine skeletons with five chiral centers, prompted us to design a method for total synthesis of 1 [3].

Many kinds of biologically active compounds containing a thiazoline moiety have been discovered or chemically synthesized. Thiazoline skeletons were generally constructed either through condensation of nitrile or imino ester with cysteamine [4] or through acid-mediated cyclization of S-acyl or N-acylcysteamine derivatives [5]. However, these thiazoline analogs were usually labile, and if they contained a chiral carbon at C-4, they were readily epimerized [6]. The application of Burgess reagent to β -hydroxythioamide is a promising method for construction of the thiazoline skeleton [7]. We have applied this method in our synthetic study. However, in the case of sterically hindered 2-(2,6-disubstituted phenyl)thiazoline as contained in micacocidin (1), this method was unsuccessful mainly due to difficulty in the preparation of the corresponding thioamide. Therefore, we searched for an effective method by which sterically hindered thiazolines could be synthesized.

Phosphorus pentachloride-mediated thiazoline construction

To find a new method for the construction of thiazolines, we found that phosphorus pentachloride (PCl₅) effected the thiazoline construction. That is, PCl₅ effectively mediated the cyclization of *N*-acyleysteamine derivatives 3 to give thiazolines 4 (**Table 1**).

$$R^{1}$$
 R^{2} R^{2} R^{2} R^{2} R^{3} R^{2} R^{4} R^{3} R^{4} R^{4} R^{4} R^{4} R^{5} R^{5} R^{2} R^{4} R^{5} R^{5

Table 1. PCl₅-mediated cyclization reactions

entry	R ¹	R ²	R ³	R ⁴	Product	yield (%)
1	Ph	Н	Н	PMB	4a	72
2	2,6-Me ₂ -Pł	n H	Н	PMB	4b	78
3	2,6-Me ₂ -Ph	n H	Н	Bn	4b	19 ^{a)}
4	2,6-Me ₂ -Pł	n H	Н	Н	4b	44 ^{b)}
5	Ph	CO ₂ Et	Н	PMB	4c	80 ^{c)}
6	Ph	CONHMe	Н	PMB	4d	trace
7	Ph	Н	Н	Bz	4a	73
8	Me ₂ CH	Н	Н	COCHMe ₂	4e	89
9	Me ₂ CH	Me	CO ₂ Me	COCHMe ₂	4f	77

a) The major product in entry 3 was a thiol (3, R^4 =H), but the reason is yet obscure.;

b) The disulfide was obtained in 20%.; c) 99.7%ee.

By this method, not only 2-alkyl- as well as 2-aryl-thiazolines, but also sterically hindered thiazolines could be prepared in good yields. Alkyl and alkoxycarbonyl were shown to be successful substituents as R^2 and R^3 in 3 (entry 5,9). However, in the case of carbamoyl (entry 6), the starting compound was decomposed. On the other hand, a *p*-methoxybenzyl (PMB) group gave favorable results as the protecting group of thiol (R^4), which was recovered as chloride S^1 after the reaction. Unprotected substrate (entry 4) resulted in dimerization of the starting thiol. Interestingly, even in the case of acyl protected thiols (entry 8, 9), the reaction proceeded smoothly and stereoselectively, and the resulting thiazolines preserved high enantiomerical purity of the starting substrate (entry 5).

Synthesis of segment A of micacocidin (1)

Our retrosynthesis of micacocidin (1) to segments A and B was as shown in **Scheme 1**, in which the labile thiazolidine ring was planned to be constructed at the latest stage possible. We considered that the C-10 chiral center, presumably generated as a mixture of the stereoisomers at the coupling reaction of segments A and B, may be converted into the desired configuration through formation of the Zn complex at the final stage. The PCl₅-mediated thiazoline construction reaction was applied to the synthesis of segment A. Synthesis of segment B (the right half of 1) and subsequent coupling with segment A will be reported in the following paper [8].

Scheme 1. Retrosynthetic analysis

^{1.} The formation of 5 was suggested from the identification of PMBCl (by ¹H-NMR and TLC) in conversion from 12 to 13.

The alkylsalicylic unit required for the construction of segment A was synthesized according to the procedure reported by Kamikawa *et al.* [9]. Wittig reaction of crotonaldehyde with phosphonium salt 8, prepared from 3-methoxy-N,N-dimethylbenzylamine (6) via benzyl chloride 7, provided a diene E/Z mixture. After hydrogenation of the diene mixture, the resulting sterically hindered ester was hydrolyzed with aqueous NaOH-DMSO under reflux to give n-pentylanisic acid 10. PCl₅-mediated cyclization of amide 12, which was prepared by condensation of 10 with protected D-cysteine 11, provided thiazoline 13 in good yield preserving high enantiomerical purity (96%ee). Another attempt to construct 13 either by condensation of a nitrile prepared from the acid 10 with cysteine or by cyclization of a thiol prepared by deprotection of 12 was unsuccessful.

NMe₂

OMe

Then CICO₂Et

OMe

To Serve 2.

NMe₂

$$n$$
-BuLi

 n -Co₂Et

 n -BuLi

 n -Co₂Ne

 n -Co₂Ne

Demethylation of both ether and ester moieties in 13 was achieved with boron tribromide to give carboxylic acid 15, which was subsequently converted to Weinreb amide 16 using bis(2-oxo-3-oxazolidinyl)phosphinic chloride (BOP-Cl), in which optical purity in the thiazoline moiety was partially lost (ca.50%ee). Phenolic ester 14 with 51%ee, initially formed after treatment of 13 with boron tribromide at low temperature, led us to presume that loss of the optical purity had already occurred at this stage. After protection of the phenol residue in 16 with t-butyldiphenylsilyl (TBDPS) group, the Weinreb amide was subjected to LiAlH₄ reduction to afford labile aldehyde 18 (protected segment A).²

^{2.} Reduction of unprotected 16 resulted in low yield, and gave extremely labile aldehyde which was not endurable for the following condensation reaction.

Without further purification, 18 was treated with N-methyl-L-cysteine hydrochloride [10] followed by desilylation with tetrabutylammmonium fluoride (TBAF) to yield thiazolidine analogs 19 as a mixture of 4 diastereomers. Treatment of the mixture with $ZnCl_2$ resulted in isomerization at the C-10 carbon into the sterically favored R configuration to provide 5-pentylpyochelin 20 (9R:9S = ca.3:1; ¹H-NMR). Comparison of ¹H-NMR data for 20 and its methyl ester 21 with those of pyochelin [11] supported the structure of 20.

In conclusion, segment A (the left half) of micacocidin (1) was efficiently synthesized using the phosphorus pentachloride-mediated thiazoline construction reaction. Successful synthesis of 5-pentylpyochelin 20 through condensation of TBDPS-Seg. A 18 with N-methyl-L-cysteine, suggested that condensation of 18 with segment B (the right half of 1) and subsequent isomerization of the C-10 chiral carbon may allow production of micacocidin (1).

Scheme 3.

Experimental section

General. Melting points were determined on a Yanagimoto micro melting point apparatus. IR spectra were recorded on a JASCO FT/IR-300E spectrometer with samples prepared as KBr pellets. 1 H-NMR spectra were recorded on a JEOL GSX-270 spectrometer. Tetramethylsilane was used as an internal standard for spectra in CDCl₃ and DMSO- d_6 . All solvents were dried over 3A or 4A molecular sieves before use. Chromatography was carried out on Merck silica gel 60. Preparative thin-layer chromatography (p-TLC) was carried out on 2.00 mm Merck silica gel $60F_{254}$ plates.

- PCl₅-mediated cyclization reaction. General procedure. To an ice-cold solution of amide 3 in CH₂Cl₂ was added PCl₅ (2.10 eq.), and the mixture was stirred at room temperature for several hours. The reaction mixture was concentrated under reduced pressure, and the residue was taken up in AcOEt and the organic extract was washed with sat.aq.NaHCO₃, brine, dried over Na₂SO₄ and then evaporated under reduced pressure. The residue was chromatographed (AcOEt/ hexane) to give thiazolines 4a-f.
- **2-Phenyl-thiazoline** (**4a**). IR v_{max} 2942, 2849, 1609, 1491, 1316, 1240, 998 cm⁻¹; ¹H-NMR (CDCl₃) δ 3.42 (2H, t, J = 8.4 Hz), 4.47 (2H, t, J = 8.4 Hz), 7.43 (3H, m), 7.84 (2H, d, J = 7.8 Hz).
- **2-(2,6-Dimetylphenyl)-thiazoline (4b).** IR v_{max} 2920, 2851, 1623, 1465, 1221, 991, 920 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.31 (6H, s), 3.48 (2H, t, J = 8.4 Hz), 4.46 (2H, t, J = 8.4 Hz), 7.03 (2H, d, J = 7.8 Hz), 7.16 (1H, t, J = 7.8 Hz).
- **4-Ethoxycarbonyl-2-phenyl-thiazoline** (**4c**). IR v_{max} 2981, 1739, 1596, 1447, 1229, 1189, 1030, 938 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.34 (3H, t, J= 7.1 Hz), 3.64 (1H, dd, J= 11.2, 9.4 Hz), 3.72 (1H, dd, J= 11.2, 8.9 Hz), 4.30 (2H, q, J= 7.1 Hz), 5.27 (1H, t, J= 9.1 Hz), 7.43 (3H, m), 7.87 (2H, d, J= 8.2 Hz).
- **2-Isopropyl-thiazoline** (**4e**). ¹H-NMR (CDCl₃) δ 1.21 (3H, s), 1.24 (3H, s), 2.79 (1H, sept, J = 6.7 Hz), 3.24 (2H, t, J = 8.5 Hz), 4.21 (2H, t, J = 8.5 Hz).
- **2-Isopropyl-4-methoxycarbonyl-4-methyl-thiazoline** (4f). ¹H-NMR (CDCl₃) δ 1.21 (3H, s), 1.23 (3H, s), 1.54 (3H, s), 2.87 (1H, sept, J = 6.7 Hz), 3.11 (1H, d, J = 11.0 Hz), 3.72 (1H, d, J = 11.0 Hz), 3.79 (3H, s).

Determination of optical purity. Optical purity of the intermediate was determined by HPLC with a chiral column [Chiralcel OD or OJ $(4.6 \times 250 \text{ mm})$, n-hexane/i-PrOH = 95/5, 1.0 ml/min, det. UV 254 or 230 nm] as shown in **Table 2**.

Table 2.

		Rt. (
Compound	Chiralcel	S isomer	R isomer	ee (%)
4a	OĎ	7.5	17.0	99.8
13	OD	9.3	7.9	96.3
14	OD	7.3	8.6	51.3
16	OJ	15.8	16.9	ca. 50

Phosphonium salt 8. To an ice-cold solution of 3-methoxy-*N*, *N*-dimethylbenzylamine (6) (25.0 g, 153 mmol) in THF (150 ml), was added *n*-BuLi (100 ml of 1.68 M in hexane, 1.10 eq.) dropwise over 10 min. The mixture was stirred at 0°C for 1 h and then cooled to -78°C, treated with ClCO₂Et (30.6 ml, 2.60 eq.), and the whole mixture was stirred further at room temperature for 20 h. The reaction mixture was diluted with AcOEt, washed with water and brine, then dried over Na₂SO₄. The solvent was removed under reduced pressure, and the residue was chromatographed (silica 1 kg, AcOEt/hexane = 1/10) to give benzyl chloride 7 (21.08 g, 60%). ¹H-NMR (CDCl₃) δ 1.40 (3H, t, J = 7.3 Hz), 3.85 (3H, s), 4.43 (2H, q, J = 7.3 Hz), 4.60 (2H, s), 6.92 (1H, d, J = 8.6 Hz), 7.03 (1H, d, J = 7.9 Hz), 7.36 (1H, dd, J = 8.6, 7.9 Hz).

A mixture of **7** (21.08 g, 92.2 mmol) and PPh₃ (24.18 g, 1.00 eq.) in toluene (92 ml) was stirred at reflux for 3 d, and then cooled to room temperature. White crystalline powders of the phosphonium salt **8** (35.83 g, 48% in 2 steps from **6**) were collected by filtration. **8**; Mp 208-211°C (decomp.); IR v_{max} 2994, 2745, 1710, 1586, 1445 cm⁻¹; ¹H-NMR (DMSO-d₆) δ 1.04 (3H, t, J = 7.3 Hz), 3.75 (3H, s), 3.99 (2H, q, J = 7.3 Hz), 4.95 (1H, s), 5.00 (1H, s), 6.56 (1H, dd, J = 7.9, 1.8 Hz), 7.12 (1H, dd, J = 8.5, 1.8 Hz), 7.30 (1H, dd, J = 8.5, 7.9 Hz), 7.55 (6H, m), 7.73 (6H, m), 7.91 (3H, m); Anal. calcd. for $C_{29}H_{28}ClO_3P$: C, 70.95; H, 5.75; Cl, 7.22. found: C, 70.84; H5.88; Cl, 7.34.

Ester 9. LDA (28.19 ml of 1.50 M in c-hexane, 1.05 eq.) was added to a suspension of phosphonium salt 8 (19.77 g, 40.3 mmol) in THF (80 ml) under a nitrogen atmosphere at -78°C over 10 min. The mixture was stirred at the same temperature for 15 min, and then at -48°C for 20 min. Crotonaldehyde (3.67 ml, 1.10 eq.) was added and the whole mixture was stirred at room temperature for 3.5 h. The reaction mixture was diluted with Et₂O and washed with brine, dried over Na₂SO₄ and then evaporated under reduced pressure. The residue was taken up in hexane, and insoluble Ph₃P=O was removed by filtration through Celite. The filtrate was concentrated under reduced pressure to yield crude diene (11.26g,

E/Z = ca.2/1).

Under a hydrogen atmosphere, a mixture of crude diene (11.26g) and 10% Pd/C (0.56g, 5wt%) in EtOH (80 ml) was stirred at room temperature for 2 d. The reaction mixture was filtered through Celite, and the filtrate was concentrated under reduced pressure. The residue was chromatographed (silica 400 g, AcOEt/hexane = 1/10) to give **9** (8.22 g, 81%). IR v_{max} 2932, 2859, 1730, 1471, 1267 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.88 (3H, t, J= 6.7 Hz), 1.32 (4H, m), 1.38 (3H, t, J= 7.3 Hz), 1.60 (2H, m), 2.55 (2H, br-t, J=7.3 Hz), 3.82 (3H, s), 4.40 (2H, q, J= 7.3 Hz), 6.75 (1H, d, J= 8.5 Hz), 6.82 (1H, d, J= 7.3 Hz), 7.26 (1H, t, J= 8.5, 7.3 Hz); LSIMS m/z 501 [2M+H]⁺, 251 [M+H]⁺, 205; HR-LSIMS m/z 251.1650 [M+H]⁺ (calcd 251.1647 for $C_{15}H_{23}O_3$).

Carboxylic acid 10. A solution of ester **9** (7.51 g, 30.0 mmol) in DMSO (45 ml) and 15% aq.NaOH (15 ml) was stirred at reflux for 15 h. After cooling to room temperature, water was added and the mixture was washed with CH_2Cl_2 . The aqueous layer was acidified with 6N aq.HCl and extracted twice with Et_2O . The combined organic extracts were washed with brine, dried over Na₂SO₄ and evaporated to give pure **10** (6.81 g, quant.). IR v_{max} 3019, 2931, 2869, 1701, 1472, 1267 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.89 (3H, t, J = 6.7 Hz), 1.34 (4H, m), 1.63 (2H, m), 2.76 (2H, td, J = 7.3 Hz), 3.90 (3H, s), 6.82 (1H, d, J = 8.5 Hz), 6.89 (1H, d, J = 7.9 Hz), 7.33 (1H, t, J = 7.9 Hz); LSIMS m/z 445 [2M+H]⁺, 223 [M+H]⁺, 205; HR-LSIMS m/z 223.1342 [M+H]⁺ (calcd 223.1334 for $C_{13}H_{18}O_3$).

Amide 12. A solution of **10**, 1,2-dichloroethane (7.5 ml), SOCl₂ (0.40 ml, 1.10 eq.) and DMF (1 drop) was stirred at reflux for 0.5 h. After cooling to room temperature, the mixture was concentrated under reduced pressure. The residue was dissolved in CH₂Cl₂ (2.3 ml), and added dropwise to a mixture of D-Cys(PMB)OMe·HCl **11** (1.60 g, 1.10 eq.), pyridine (1.01 ml, 2.50 eq.) and CH₂Cl₂ (6.0 ml) over 10 min at 0°C. The mixture was stirred at room temperature for 1 h, and concentrated to dryness. The residue was taken up in AcOEt and the organic extract was washed with sat.aq.NH₄Cl, sat.aq.NaHCO₃ and brine, dried over Na₂SO₄ and then evaporated under reduced pressure. The residue was chromatographed (silica 100 g, AcOEt/hexane = 1/2) to give **12** (2.18 g, 95%). Mp 71-73 °C; $[\alpha]_{D}^{24}$ +5.0 (c 1.01, CHCl₃); IR v_{max} 3356, 2951, 2855, 1745, 1662, 1512, 1247 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.87 (3H, t, J = 6.7 Hz), 1.30 (4H, m), 1.60 (2H, m), 2.65 (2H, br-t, J =7.3 Hz), 2.93 (1H, dd, J = 14.0, 4.9 Hz), 3.06 (1H, dd, J = 14.0, 4.9 Hz), 3.72 (2H, s), 3.78 (9H, s), 5.07 (1H, dt, J = 7.9, 4.9 Hz), 6.58 (1H, br-d, J = 7.9 Hz), 6.75 (1H, d, J = 8.5 Hz), 6.83 (3H, m), 7.24 (3H, m); Anal. calcd for C₂₅H₃₃NO₅S: C, 65.33; H, 7.24; N, 3.05. found: C, 65.20; H, 7.21; N,3.09.

Thiazoline ester 13. Following treatment similar to the general procedure, amide **12** (2.07 g, 4.50 mmol) gave **13** (1.25 g, 86%). $[\alpha]_D^{27}$ -29.1 (c 1.00, CHCl₃); IR ν_{max} 2953, 2858, 1745, 1469, 1265 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.88 (3H, t, J = 6.7 Hz), 1.30 (4H, m), 1.59 (2H,

m), 2.61 (2H, br-t, J= 7.3 Hz), 3.69 (1H, dd, J= 11.0, 9.2 Hz), 3.77 (1H, dd, J= 11.0, 9.2 Hz), 3.80 (3H, s), 3.84 (3H, s), 5.30 (1H, t, J= 9.2 Hz), 6.74 (1H, d, J= 8.5 Hz), 6.83 (1H, d, J= 7.3 Hz), 7.26 (1H, t, J= 8.5, 7.3 Hz); LSIMS m/z 643 [2M+H]⁺, 322 [M+H]⁺, 262; HR-LSIMS m/z 322.1468 [M+H]⁺ (calcd 322.1477 for $C_{17}H_{24}NO_3S$).

Weinreb amide 16. To a solution of thiazoline ester 13 (1.13 g, 3.52 mmol) in CH₂Cl₂ (7.0 ml) was added BBr₃ (7.38 ml of 1.00 M in CH₂Cl₂, 2.10 eq.) at -78°C. The mixture was stirred at the same temperature for 15 min, and then at room temperature for a further 3.5 h. Pieces of ice were added, and the whole mixture was diluted with AcOEt. The organic mixture was washed with water, brine, then dried over Na₂SO₄ and concentrated under reduced pressure to give acid 15 (0.73 g, 71%). IR v_{max} 3031, 2929, 2857, 1730, 1598, 1452 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.90 (3H, t, J= 6.7 Hz), 1.35 (4H, m), 1.60 (2H, m), 2.92 (2H, m), 3.69 (2H, m), 4.29 (2H, br-s), 5.37 (1H, t, J= 9.2 Hz), 6.73 (1H, d, J= 8.5 Hz), 6.86 (1H, d, J= 7.9 Hz), 7.23 (1H, t, J= 8.5, 7.9 Hz).

A mixture of carboxylic acid **15** (686 mg, 2.34 mmol), MeONHMe·HCl (274 mmol, 1.20 eq.), BOP-Cl (655 mg, 1.10 eq.), CH₂Cl₂ (7.0 ml) and Et₃N (0.82 ml, 2.50 eq.) was stirred at room temperature overnight, and then concentrated. The residue was taken up in AcOEt and the organic extract was washed with 5% aq.KHSO₄, sat.aq.NaHCO₃ and brine, dried over Na₂SO₄ and then evaporated under reduced pressure. The residue was chromatographed (silica 30 g, AcOEt/hexane = 1/2) to give Weinreb amide **16** (539 mg, 48% calculated from **13**). IR ν_{max} 3292, 2931, 2856, 1653, 1582, 1453, 1208 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.90 (3H, t, J = 6.7 Hz), 1.35 (4H, m), 1.63 (2H, m), 2.90 (1H, m), 3.30 (3H, br-s), 3.55 (1H, dd, J = 11.0, 9.8 Hz), 3.70 (2H, m), 3.84 (3H, s), 5.65 (1H, br-t, J = 7.3 Hz), 6.72 (1H, d, J = 7.9 Hz), 6.84 (1H, d, J = 7.9 Hz), 7.21 (1H, t, J = 7.9 Hz)); LSIMS m/z 673 [2M+H]⁺, 337 [M+H]⁺, 248; HR-LSIMS m/z 337.1584 [M+H]⁺ (calcd 337.1586 for $C_{17}H_{25}N_2O_3S$).

TBDPS ether 17. A mixture of Weinreb amide 16 (200 mg, 0.59 mmol), DMF (1.20 ml), TBDPSCl (343 mg, 2.10 eq.) and imidazole (170 mg, 4.20 eq.) was stirred at 50 °C for 2 h. After cooling to room temperature, the mixture was diluted with Et₂O and washed with water, brine, then dried over Na₂SO₄ and concentrated under reduced pressure. The residue was chromatographed (silica 20 g, AcOEt/hexane = 1/6) to give 17 (315 mg, 92%). IR ν_{max} 2931, 2859, 1670, 1463, 1283 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.89 (3H, t, J = 6.7 Hz), 1.06 (9H, s), 1.33 (4H, m), 1.62 (2H, m), 2.63 (2H, br-t, J = 7.3 Hz), 3.30 (3H, br-s), 3.52 (1H, dd, J = 11.0, 9.2 Hz), 3.82 (3H, s), 4.01 (1H, br-t, J = 9.2 Hz), 5.67 (1H, br-t, J = 9.2 Hz), 6.21 (1H, d, J = 7.9 Hz), 6.72 (1H, d, J = 6.7 Hz), 6.82 (1H, t, J = 7.9, 6.7 Hz), 7.37 (6H, m), 7.74 (4H, m); LSIMS m/z 575 [M+H]⁺, 517; HR-LSIMS m/z 575.2766 [M+H]⁺ (calcd 575.2764 for $C_{33}H_{43}N_2O_3SSi$).

5-Pentylpyochelin 20. To an ice-cold solution of TBDPS ether **17** (75.0 mg, 0.13 mmol) in THF (2.00 ml) was added LiAlH₄ (5.40 mg, 1.10 eq.), and the mixture was stirred at the same temperature for 30 min. The reaction was quenched with AcOEt, then the mixture was poured into sat.aq.NH₄Cl and extracted with AcOEt. The AcOEt layer was washed with water and brine, dried over Na₂SO₄, and concentrated in vacuo to give crude aldehyde **18** as a bright yellow amorphous solid.

Without further purification, 18 was dissolved in CH₂Cl₂ (4.00 ml) and MeOH (1.0 ml), then AcOK (128 mg, 10.0 eq.) and N-Me-L-Cys•HCl (ca. 50% purity, 116 mg, 2.50 eq.) were added to the solution, and the mixture was stirred at room temperature overnight. The reaction mixture was diluted with AcOEt, washed with 5% aq.KHSO₄ and brine, dried over Na₂SO₄, and then concentrated in vacuo.

To an ice-cold solution of the residue in THF (3.00 ml) was added TBAF (1.00 M in THF, 0.13 ml, 1.00 eq.), and the mixture was stirred at room temperature for 30 min. The reaction mixture was diluted with AcOEt and washed with 5% aq.KHSO₄, then extracted into sat.aq.NaHCO₃ The combined aqueous layer was acidified (pH=4) with conc.HCl and extracted again with AcOEt. The AcOEt layer was washed with brine, dried over Na₂SO₄, then concentrated in vacuo to give 5-pentylpyochelin **19** (37.0 mg) as a mixture of diastereomers.

To a solution of the residue (37.0 mg, 9.37 x 10^{-5} mol) in MeOH (2.50 ml), was added ZnCl₂ (32.0 mg, 2.5 eq.), and the whole mixture was stirred at room temperature overnight. The reaction mixture was diluted with AcOEt and washed with 5% aq.KHSO₄ to release the zinc ion. The organic layer was washed with brine, dried over Na₂SO₄, and then concentrated in vacuo to give 5-pentylpyochelin **20** (ca.3:1 mixture of diastereomers; ¹H-NMR, 37.0 mg) as yellow caramel. IR v_{max} 3014, 2930, 2859, 1725, 1581, 1464, 1209 cm⁻¹; ¹H-NMR (CDCl₃) [major isomer **20**] δ 0.90 (3H, t, J = 6.8 Hz), 1.35 (4H, m), 1.62 (2H, m), 2.72 (3H, s), 2.95 (2H, m), 3.21 (1H, dd, J = 11.4, 7.8 Hz), 3.29-3.54 (3H, m), 3.90 (1H, dd, J = 7.4, 5.8 Hz), 4.33 (1H, d, J = 8.6 Hz), 4.70 (1H, q, J = 8.6 Hz), 6.73 (1H, d, J = 6.9 Hz), 6.87 (1H, d, J = 7.2 Hz), 7.24 (1H, t, J = 7.2, 6.9 Hz).

5-Pentylpyochelin methyl ester 21. A solution of **20** (37.0 mg) in CH₂Cl₂ (3.00 ml) and MeOH (1.00 ml) was treated with TMSCHN₂ (1.00M in THF) until generation of N₂ gas ceased. The reaction was quenched with AcOH and the mixture was concentrated in vacuo. The residue was purified by p-TLC (AcOEt/hexane = 1/1) to give 5-pentylpyochelin methyl ester **21** (ca.3:1 mixture of diastereomers by ¹H-NMR, 24.0 mg, 45% from **17**). IR v_{max} 2955, 2870, 2716, 1739, 1581, 1447, 1208 cm⁻¹; ¹H-NMR (CDCl₃) [major isomer; presume 9*R*] δ 0.90 (3H, t, J = 6.9 Hz), 1.36 (4H, m), 1.62 (2H, m), 2.59 (3H, s), 2.97 (2H, t, J = 7.9 Hz), 3.13 (1H, dd, J = 10.7, 6.4 Hz), 3.23 (1H, dd, J = 10.7, 8.6 Hz), 3.41 (2H, m), 3.68 (1H, dd, J = 8.6, 6.4 Hz), 3.76 (3H, s), 4.50 (1H, d, J = 5.6 Hz), 4.94 (1H, td, J = 8.7, 5.6 Hz), 6.69 (1H,

dd, J = 7.6, 1.2 Hz), 6.84 (1H, dd, J = 8.2, 1.2 Hz), 7.19 (1H, t, J = 7.8 Hz); [minor isomer; presume 9S] δ 0.90 (3H, t, J = 6.9 Hz), 1.36 (4H, m), 1.62 (2H, m), 2.60 (3H, s), 2.97 (2H, t, J = 7.9 Hz), 3.20-3.57 (4H, m), 3.77 (3H, s), 3.82 (1H, t, J = 6.8 Hz), 4.23 (1H, d, J = 7.9 Hz), 4.69 (1H, q, J = 7.9 Hz), 6.69 (1H, d, J = 7.6 Hz), 6.84 (1H, d, J = 8.2 Hz), 7.19 (1H, t, J = 7.8 Hz); LSIMS m/z 409 [M+H]⁺, 248, 160; HR-LSIMS m/z 409.1612 [M+H]⁺ (calcd 409.1620 for $C_{20}H_{20}N_2O_3S_2$).

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References

- [1] a) Lemaitre, M.; Guetard, D.; Henin, Y.; Montagnier, L.; Zerial, A. Res. Viol. 1990, 141, 5-16. b) Lo, S-C.; Dawson, M.S.; Wong, D.M.; Newtoon III, P.B.; Sonoda, M.A.; Engler, W.F.; Wang, R.Y.H.; Shin, J.W.K; Alter, H.J.; Wear, D.J. Am. J. Trop. Med. Hyg. 1989, 41, 601-616. c) Chowdhury, M.I.H.; Munakata, T.; Koyanagi, Y.; Kobayashi, S.; Arai, S.; Yamamoto, N. Biochem. Biophys. Res. Commun. 1990, 170, 1365-1370. d) Katseni, V.L.; Gilroy, C.B.; Ryait, B.K.; Ariyoshi, K.; Bieniasz, P.D.; Weber, J.N.; Robinson, D.T. Lancet 1993, 341, 271-273.
- [2] a) Shionogi & Co., Ltd. Patent, WO 96/04262. b) Kobayashi, S.; Hidaka, S.; Kawamura, Y.; Ozaki, M.; Hayase, Y. J. Antibiotics 1998, 51, 323-327. c) Kobayashi, S.; Nakai, H.; Ikenishi, Y.; Sun, Y.; Ozaki, M.; Hayase, Y.; Takeda, R. ibid. 1998, 51, 328-332.
- [3] Ino, A.; Hasegawa, Y.; Murabayashi, A. Tetrahedron Lett. 1998, 39, 3509-3512.
- [4] a) Mulqueen, G.C.; Pattenden, G.; Whiting, D.A. Tetrahedron 1993, 49, 5359-5364. b) Boyce, R.J.; Pattenden, G. ibid. 1995, 51, 7313-7320. c) Boyce, R.J.; Mulqueen, G.C.; Pattenden, G. ibid. 1995, 51, 7321-7330. d) Rinehart, K.L.; Staley, A.L.; Wilson, S.R.; Ankenbauer, R.G.; Cox, C.D. J. Org, Chem. 1995, 60, 2786-2791. e) Kwiatkowski, S.; Crocker, P.J.; Chavan, A.J.; Imai, N.; Haley, B.E.; Watt, D.S.; Ho, R.-J. Tetrahedron Lett. 1990, 31, 2093-2096. f) Toya, Y.; Takagi, M.; Kondo, T.; Nakata, H.; Isobe, M.; Goto, T. Bull. Chem. Soc. Jpn. 1992, 65, 2604-2610. g) Caujolle, R.; Favrot, J.D.; Loiseau, P.R.; Payard, M.; Amarouch, H.; Lazrek, H.; Linas, M.D.; Segueda, J.P.; Loiseau, P.M.; Bories, C.; Gayral, P. Pharm. Acta. Helv. 1991, 66, 237-240. h) Crawhall, J.C.; Elliott, D.F.; Hooper, K.C. J. Chem. Soc. 1956, 4066.
- [5] a) White, J.D.; Kim, T.-S.; Nambu, M. J. Am. Chem. Soc. 1997, 119, 103-111. b) Fukuyama, T.; Xu, L. J. Am. Chem. Soc. 1993, 115, 8449-8450. c) Wipf, P.; Venkatraman, S. J. Org, Chem. 1995, 60, 7224-7229. d) Walker, M.A.; Heathcock, C.H. ibid. 1992, 57, 5566-5568. e) Akaji, K.; Kuriyama, N.; Kiso, Y. ibid. 1996, 61, 3350-3357. f) Onoda, T.; Shirai, R.; Koiso, Y.; Iwasaki, S. Tetrahedron 1996, 52, 14543-14562.
- [6] a) Wipf, P.; Fritch, P.C. J. Am. Chem. Soc. 1996, 118, 12358-12367. b) Wipf, P.; Fritch, P.C. Tetrahedron Lett. 1994, 35, 5397-5400.

- [7] a) Wipf, P.; Miller, C.P. Tetrahedron Lett. 1992, 33, 907-910. b) Wipf, P.; Miller, C.P.; Venkatraman,
 S.; Fritch, P.C. Tetrahedron Lett. 1995, 36, 6395-6398. c) Wipf, P.; Xu, W. J. Org, Chem. 1996, 61, 6556-6562.
- [8] Ino, A.; Hasegawa, Y.; Murabayashi, A. Tetrahedron, following paper.
- [9] Kubo, I.; Kim, M.; Naya, K.; Komatsu, S.; Yamagiwa, Y.; Ohashi, K.; Sakamoto, Y.; Hirakawa, S.; Kamikawa, T. Chem. Lett. 1987, 1101-1104.
- [10] Blondeau, P.; Berse, C.; Gravel, D. Can. J. Chem. 1967, 45, 49.
- [11] a) Cox, C.D.; Rinehart, K.L., Jr.; Moore, M.L.; Cook, J.C., Jr. Proc. Natl. Acad. Sci. U.S.A. 1981, 78, 4256-4260. b) Ankenbauer, R.G.; Toyokuni, T.; Staley, A.; Rinehart, K.L., Jr.; Cox, C.D. J. Bacteriol. 1988, 170, 5344-5351. c) Rinehart, K.L.; Staley, A.L.; Wilson, S.R.; Ankenbauer, R.G.; Cox, C.D. J. Org, Chem. 1995, 60, 2786-2791.